

Eczema and allergy: the chicken or the egg ?

Aurélie Collins, Bénédicte Derkenne

CHR Verviers, Department of Paediatric Allergy, Verviers, Belgium

Aurelie.collins@chrverviers.be

Keywords

Eczema ; atopic dermatitis ; allergy.

Abstract

Eczema is a common problem in general paediatric consultations. Treatment of eczema is essential because irritated and inflamed skin stimulates the onset of sensitisation and subsequent allergy, particularly food allergy. Treatment is currently based on non-aggressive corporal hygiene, daily application of emollients and early treatment of flares with topical corticosteroids.

In severe cases, an allergic aetiology is often suspected. Early-onset eczema before the age of 6 months and/or steroid-dependent eczema are more likely to be allergic in origin. In these cases, cow's milk is the main culprit, either in artificial milk or via breast milk. Eviction would help to reduce the intensity of the disease. In the older children, an allergic trigger is found in only 10% of cases.

However, a very large number of patients have asymptomatic sensitisation (positive skin prick test and/or positive specific IgE) which should not be confused with allergy. In fact, if eliminating the food does not improve the cutaneous symptoms, it is not only useless but may even promote a breach of tolerance and increase the risk of becoming truly allergic to that food.

Introduction

Atopic dermatitis (AD) is the most common chronic inflammatory skin disease in children. In its moderate to severe form, it has a significant impact on quality of life, for example through sleep disturbance, but also because of the cost and time involved in treatment.

The prevalence of AD is difficult to estimate because of the different terms are used to describe it. It is to be between 10 and 20% in the paediatric population, according to different studies, and has been increasing steadily in recent years. This increase is mainly observed in developing countries where the prevalence is low, with a stabilisation in countries where the prevalence is already high (1,2).

AD : a multifactorial disease

There is undoubtedly a genetic predisposition. Indeed, about 70% of patients with AD have a positive family history of atopy, and the odds ratio for developing AD is 2-3 times higher in children with 1 atopic parent and 3-5 times higher if both parents are atopic (3).

Genetic mutations explain this trend, including loss-of-function mutations in filaggrin (FLG), a structural protein of the epidermis, but also mutations in the lipidic cement, keratinocytes and antimicrobial peptides. These mutations lead to a dysfunction of the skin barrier, allowing water to escape to the outside, leading to a skin xerosis and allowing the penetration of pathogenic microorganisms and allergens into the skin tissue, promoting superinfections and the development of secondary allergic sensitisation.

There is also a dysfunction of the immune response, with an imbalance between the Th1/Th2 response, promoting the IgE-mediated response, on the one hand, and the secretion of cytokines self-sustaining the capacity of the immune response on the other. Furthermore, an involvement of the *Staphylococcus aureus* superantigen has also been demonstrated.

However, the recent increase of AD as well as of other atopic diseases is too rapid to be explained by genetics alone. So, there is an involvement of the environment. In this sense, the hygiene hypothesis suggests that

the transition from a rural to an urban lifestyle has reduced children's exposure to some pathogenic microorganisms, and consequently altered the Th1-Th2 balance. By limiting repeated exposure to microbes, the Th1 response is less provoked, and the Th2 response is more promoted inducing atopic manifestations in infants and young children living in a more sterile environment. Other environmental factors have also been taken into account: breastfeeding, changes in dietary habits (obesity), smoking, exposure to domestic furry animals, exposure to dust mites and cockroaches, overheating of homes, pollutants and climatic factors, lack of exposure to UVB, use of alkaline detergents, perfumes and preservatives (4 - 7).

AD: diagnosis and treatment

The diagnosis of AD is clinical. The UK Working Party diagnostic criteria for atopic dermatitis were established in 1994 and have been widely used in studies and are easy to use in everyday practice.

A history of itchy skin plus at least 3 of the following criteria :

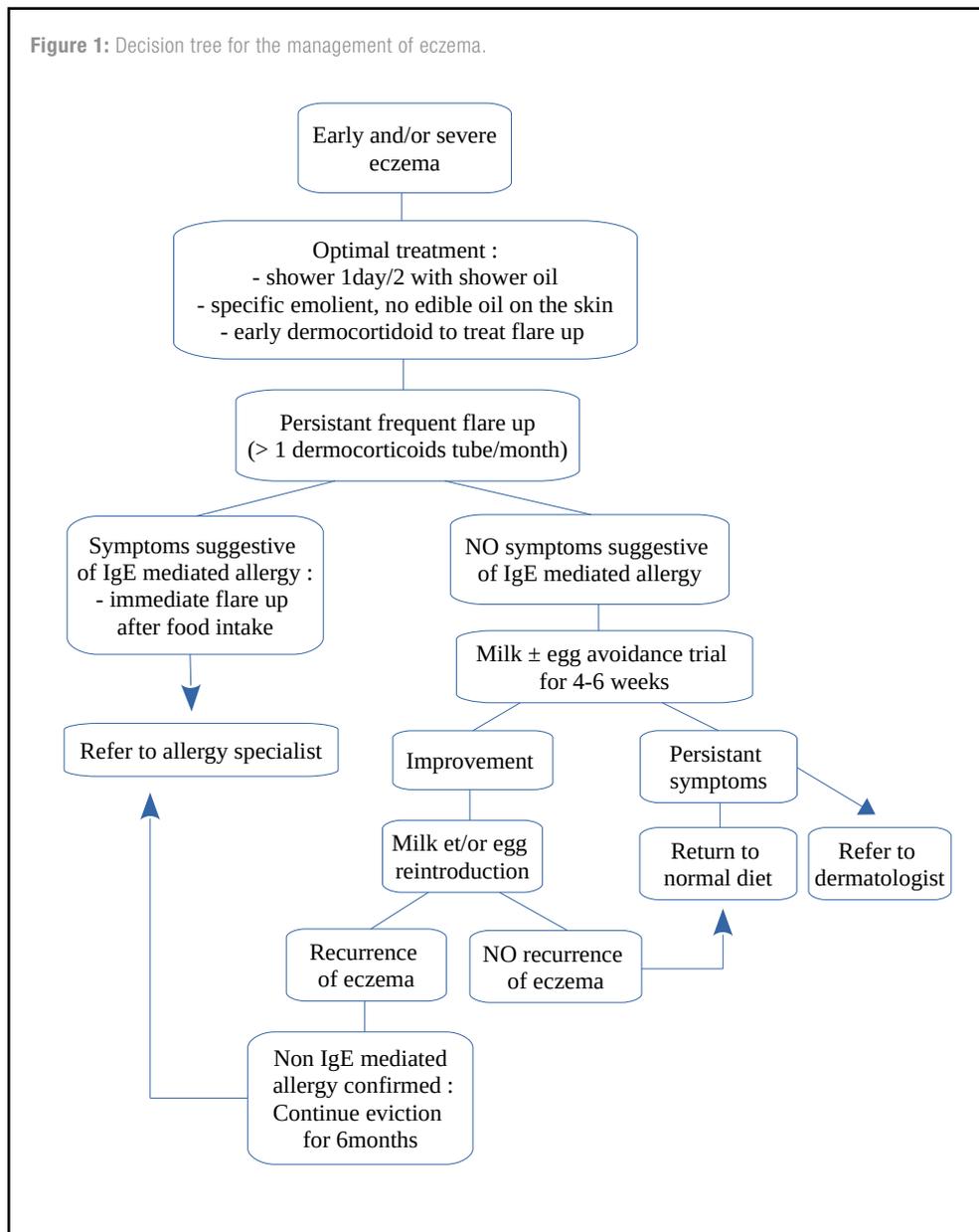
- visible flexural dermatitis involving the skin creases, such as the bends of the elbows
- personal history of flexural dermatitis (or dermatitis on the cheeks and/or extensor areas in children aged 18 months or younger)
- personal history of dry skin in the past 12 months
- personal history of asthma or allergic rhinitis (or history of atopic disease in a first-degree relative of children aged less than four years)
- onset of signs and symptoms before the age of two years.

There are also several severity scores (EASI: eczema area and severity index, SCORAD: severity scoring atopic dermatitis index, POEM: patient-oriented eczema measure), but these are cumbersome and not always very useful in current practice.

The cornerstone of treatment is local care. Hygiene measures to strengthen the skin barrier (short warm baths 1-2 times a week, frequent daily application of cutaneous emollients) and regulation of the immune response by early anti-inflammatory treatment (with dermocorticoids and calcineurin inhibitors, the latter only reimbursed in Belgium from the age of 2 years).

There are two approaches: reactive treatment of flare-ups and proactive treatment even outside flare-ups to limit them. The ongoing PACI study is an RCT that aims to determine the best approach (8).

A decision tree for the management of eczema is shown in Figure 1.



AD and allergic triggers

In case of severe eczema, an allergic trigger is often sought. A thorough medical history is essential. In fact, systematic testing is not recommended. A suggestive history of food allergy or a moderate to severe eczema with little or no response to optimal topical treatment should raise suspicion of an underlying allergy. On the other hand, unwarranted investigation leads to many false positive reactions. Indeed, there is a high prevalence of asymptomatic food sensitisations in AD (9). Misinterpretation of test results (specific IgE's or prick tests) can lead to severe elimination diets with negative effects on quality of life, but also to avoidance or postponement of tolerance and even induction of food allergy (10).

The EAACI (European Academy of Allergy and Clinical Immunology) recommends testing for food allergens in case of eczema in the following situations (11) :

- AD and immediate reaction to one or more foods
- Persistent, moderate to severe AD with no history of immediate reaction to a food
- Foods suspected by the patient or his family with no obvious history of immediate reaction.

Skin prick tests (SPT) and specific IgE's have a good negative predictive value of 95%, but a bad positive predictive value of only 50%. Atopy patch tests are more sensitive but less specific. However, they are not recommended by EAACI due to their lack of standardisation.

Depending on the equipment available, one test or the other may be preferred. The SPT, which consists of pricking the skin with a lancet through a drop of a commercially available allergen extract or fresh food, is usually preferred because of its low cost, speed of execution and interpretation, and the ability to test for multiple allergens at once. On the other hand, they are generally preferred by children over blood testing. However, in general practice, specific IgE's are often preferred for practical reasons due to lack of experience and/or appropriate equipment.

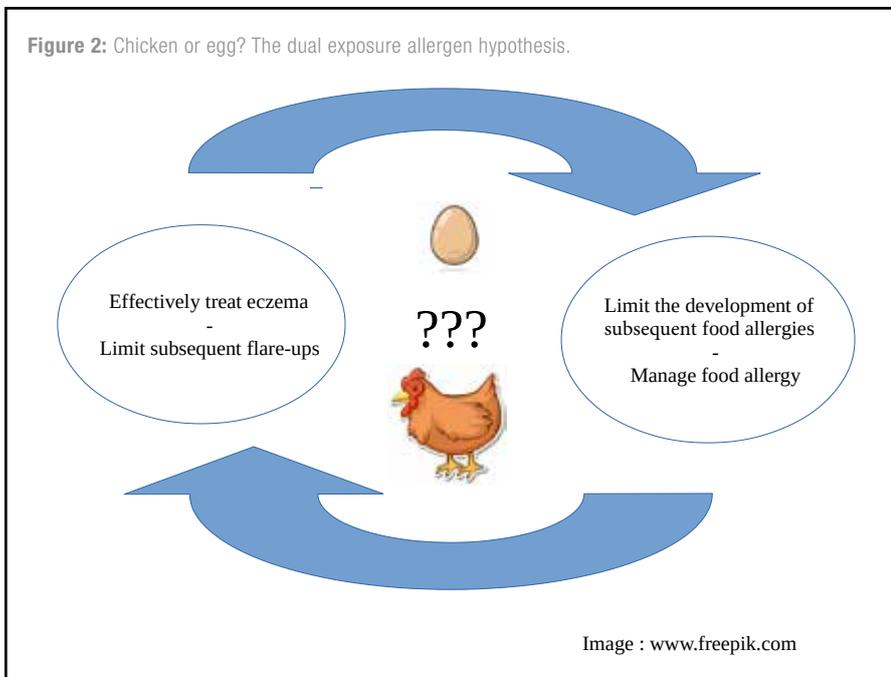
If a food trigger is suspected, the food challenge remains the gold standard to confirm the allergy before setting up an elimination period or after a trial elimination diet. In the case of eczema, an elimination period of 4 to 6 weeks followed by a reintroduction to confirm the food's involvement in the symptomatology is essential. In the specific case of a positive test for a food that has not yet been introduced into the child's diet, caution is required as there is a risk of real allergy due to allergic sensitisation and the first introduction must be made under medical supervision.

Especially in case of early onset eczema (<6 months of life) and / or corticosteroid-dependent eczema, it is more likely to act as an allergic trigger. In this case, it is mainly cow's milk, either in formula or via breast milk, but it can also be egg or peanut (12). Elimination of the food would help to reduce the severity of the disease. In older children, an allergy is found in only 10% of cases.

AD and subsequent food allergy

In 2008, a new hypothesis on the aetiology of food allergy emerged (the dual exposure allergen hypothesis). The site of a food's first encounter with the immune system determines the subsequent tolerance or allergy to that food (Figure 2).

Figure 2: Chicken or egg? The dual exposure allergen hypothesis.



The digestive tract is mainly tolerogenic, whereas the cutaneous route is more likely to lead to allergy (13). Since then, several other elements have complicated this theory, e.g. implication of vitamin D levels, skin flora (14).

In children carrying one or more loss-of-function mutations in the filaggrin gene, a dose-response relationship has been demonstrated between early life environmental exposure to peanut protein in house dust and subsequent sensitisation and allergy to peanuts (15).

We can therefore understand that early introduction of foods promotes their tolerance and that a delayed introduction increases the risk of allergy. The latest recommendations are to introduce the main allergens (egg and peanut) as early as 4-6 months.

Similarly, it is recommended that potential allergens should not be applied to weakened and damaged skin. We should avoid creams made with food proteins and prefer creams containing few ingredients (16).

Treatment of eczema should also be started early to reduce the risk of developing a subsequent food allergy. However, it remains a major challenge because of the significant corticophobia, of both the family and some of the patient's health care providers (physicians, paramedics, pharmacists ,...) (17).

Conclusion

Take home messages

- AD is a common skin disease in children with a significant impact on quality of life.
- The tolerogenic role of gastrointestinal exposure versus cutaneous exposure needs to be recognised.
- Restoring the cutaneous barrier is very important.
- Fighting corticophobia is one of our tasks.
- Early introduction of allergenic foods (egg and peanut between 4-6months) must be advocated.
- Advise food avoidance only when justified by history, allergy testing (specific IgE and prick testing) and/or oral food challenge..
- Referral to an allergologist may be necessary for severe AD not controlled by dermatocorticoids.

Conflicts of interest

The authors have no conflicts of interest to declare with regard to the topic discussed in this manuscript.

REFERENCES

1. Asher MI, Montefort S, Björkstén B, Lai CKW, Strachan DP, Weiland SK, et al. ISAAC Phase Three Study Group. Worldwide time trends in the prevalence of symptoms of asthma, allergic rhinoconjunctivitis, and eczema in childhood: ISAAC Phases One and Three repeat multicountry cross-sectional surveys. *Lancet*. 2006;368(9537):733-43.
2. Deckers IAG, McLean S, Linssen S, Mommers M, van Schayck CP, Sheikh A. Investigating international time trends in the incidence and prevalence of atopic eczema 1990-2010 : a systematic review of epidemiological studies. *Plos One*. 2012; 7(7):e39803.
3. Böhme M, Wickman M, Lennart Nordvall S, Svartengren M, Wahlgren CF. Family history and risk of atopic dermatitis in children up to 4 years. *Clin Exp Allergy*. 2003;33(9):1226-31.
4. Kathuria P, Silverberg JI, Association of pollution and climate with atopic eczema in US children. *Pediatr Allergy Immunol*. 2016;27(5):478-85.
5. Thyssen JP, Zirwas MJ, Elias PM, Potential role of reduced environmental UV exposure as a driver of the current epidemic of atopic dermatitis. *J Allergy Clin Immunol*. 2015;136(5):1163-9.
6. Boothe WD, Tarbox JA, Tarbox MB, Atopic Dermatitis: Pathophysiology. *Adv Exp Med Biol*. 2017;1027:21-37.
7. Sozener ZC, Yucel UO, Altiner S, Ozturk BO, Cerci P, Turk M, et al. The external exposure and allergies: from the perspective of the epithelial barrier hypothesis. *Front Allergy*. 2022;3:887672.
8. Yamamoto-Hanada K, Kobayashi T, Williams HC, Mikami M, Saito-Abe M, Morita K, et al. Early aggressive intervention for infantile atopic dermatitis to prevent development of food allergy: a multicenter, investigator-blinded, randomized, parallel group controlled trial (PACI Study)-protocol for a randomized controlled trial. *Clin Transl Allergy*. 2018;23;8:47.
9. Robison R, Singh AM, Controversies in allergy : food testing and dietary avoidance in atopic dermatitis. *J Allergy Clin Immunol Pract*. 2019;7(1):35-9.
10. Chang A, Robison R, Cai M, Singh AM, Natural history of food-triggered atopic dermatitis and development of immediate reactions in children. *J Allergy Clin Immunol Pract*. 2016;4(2):229-36.e1.
11. Werfel T, Ballmer-Weber B, Eigenmann PA, Niggemann B, Rancé F, Turjanmaa K, et al. Eczematous reactions to food in atopic eczema : position paper of the EAACI and GA2LEN. *Allergy*. 2007;62(7):723-8.
12. Eigenmann PA, Calza AM, Diagnosis of IgE-mediated food allergy among swiss children with atop ermatitistis. *Pediatr Allergy Immunol*. 2000;11(2):95-100.
13. Lack G, Epidemiologic risks for food allergy. *J Allergy Clin Immunol*. 2008;121:1331-6.
14. Du Toit G, Sampson HA, Plaut M, Burks AW, Akdis CA, Lack G, Food allergy: Update on prevention and tolerance. *J Allergy Clin Immunol*. 2018;141(1):30-40.
15. Brough HA, Simpson A, Makinson K, Hankinson J, Brown S, Douiri A, et al. Peanut allergy: effect of environmental peanut exposure in children with filaggrin loss-of-function mutations. *J Allergy Clin Immunol*. 2014;134(4):867-875.
16. Lack G, Fox D, Northstone K, Golding J. Avon longitudinal study of parents and children study team. Factors associated with the development of peanut allergy in childhood. *N Engl J Med*. 2003;348(11):977-85.
17. Brough HA, Nadeau KC, Sindher SB, Alkotob SS, Chan S, Banhson HT, et al. Epicutaneous sensitization in the development of food allergy: What is the evidence and how can this be prevented? *Allergy* .2020;75(9):2185-205.

Drug hypersensitivity reactions: an overview

Athina L van Gasse ^{a,c}, Marie-Line M van der Poorten ^{a,c}, Margo M Hagendorens ^{a,c}, Vito Sabato ^{a,b}, Didier G Ebo ^{a,b}

^a Faculty of Medicine and Health Sciences, Department of Immunology, Allergology, Rheumatology and the Infla-Med Centre of Excellence, University of Antwerp and Antwerp University Hospital, Antwerpen (Belgium)

^b AZ Jan Palfijn Gent, Department of Immunology and Allergology, Ghent (Belgium)

^c Faculty of Medicine and Health Sciences, Department of Paediatrics and the Infla-Med Centre of Excellence, University of Antwerp, and Antwerp University Hospital, Antwerpen (Belgium)

Didier Ebo: immuno@uantwerpen.be

Keywords

Drug hypersensitivity; hypersensitivity; immunoglobulin E; T-lymphocytes; penicillins; diagnostic errors.

Abstract

Drug hypersensitivity reactions (DHRs) account for 10% of all adverse drug reactions.

DHRs are clinically classified as immediate, mostly drug-specific IgE antibody (sIgE) -mediated, and nonimmediate, mostly T-cell mediated, reactions. Gaining insights into the underlying pathophysiological mechanism is crucial for correct orientation of further diagnostic work-up of DHRs. Therefore, a thorough history focusing on elements such as signs, symptoms, timing, index drug, re-exposition is of paramount importance. In case of immediate DHR, diagnosis may comprise skin testing with immediate readings, sIgE antibody quantification, specialized *in vitro* diagnostics. In nonimmediate DHR, sIgE antibodies are not useful and skin tests are performed with delayed readings. In difficult cases with negative or uncertain test results, eventually a drug challenge might be required to document or refute diagnosis.

Correct diagnosis of DHRs is very important. Unverified and false diagnoses of “drug allergy”, mainly “penicillin allergy”, have evolved into a plague with increasing medical and financial burden. On the other hand, misdiagnosis entails a risk for potentially life-threatening and fatal reactions upon re-exposure. Therefore, quick referral for an allergy workup in case of a possible DHR is recommended.

Introduction

Adverse drug reactions (ADRs) are defined as unintended, harmful effects resulting from exposure to a compound for diagnostic, prophylactic, or therapeutic purposes. Most ADRs directly dependent on the pharmacological properties of the drug (e.g. bleeding by anti-coagulants). Drug hypersensitivity reactions (DHRs) on the other hand, comprise symptoms resulting from effects extending beyond the pharmacological targets of a drug and can result from the activation of immune cells, inflammatory pathways, or both. DHRs account for 10% of all ADRs. According to the World Allergy Organization (WAO), DHRs occur in 1% to 2% of all admissions and in 3% to 5% of the hospitalized patients. The true prevalence in the community is unknown. However, despite absence of correct prevalence data in children, DHR are estimated to be less frequent than in adults, possibly because of less exposure to drugs.

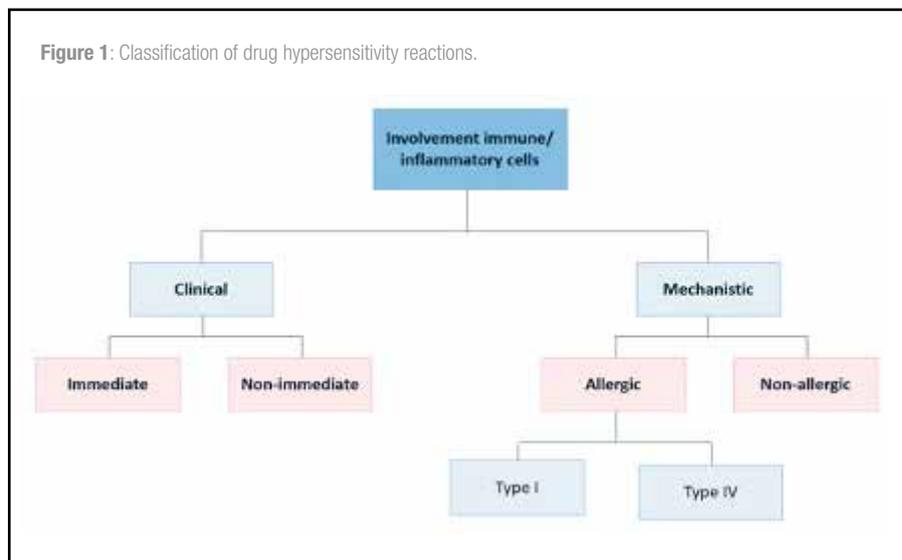
As shown in *Figure 1*, DHRs can be classified based upon their clinical presentation (chronology and morphology) and/or based upon their underlying pathophysiological mechanism.

According to the underlying pathophysiological mechanism, DHRs can be further subclassified as allergic or non-allergic. Allergic hypersensitivity involves specific activation

of drug-reactive T- and/or B-cells of the adaptive immune system (1). T-cells are involved in both immediate and nonimmediate allergic reactions. In contrast, B-cells are only involved in IgE-mediated immediate allergic reactions. Traditionally, 4 types of allergic DHRs are described according to the Gell and Coombs classification (table 1) (2). Type I (mostly mediated by drug-specific IgE antibodies (sIgE)) and type IV (cell mediated) reactions are the most frequent encountered reactions.

In contrast, non-allergic immediate DHRs (IDHRs) involve activation of

Figure 1: Classification of drug hypersensitivity reactions.



immune cells and release of mediators by direct mechanisms independent from the adaptive immune system response (e.g., mast cell activation via activation of the mas-related G protein-coupled receptor type X2 (MRGPRX2) or due to pro-inflammatory mediators increased by COX-1 inhibition). Established MRGPRX2-agonists are opiates, quinolones, and neuromuscular blocking agents (3, 4, 5). However, these drugs can also trigger sIgE-dependent basophil and mast cell degranulation. Non-allergic nonimmediate DHRs (NIDHRs) can also result from pharmacological interaction between a drug and MHC of the antigen-presenting cell or T-cell receptor (6).

As shown in figure 1, from a clinical point of view, DHRs are clinically classified as immediate and nonimmediate reactions, designated respectively as IDHRs and NIDHRs. Immediate reactions usually occur within 1 hour, also depending on the route of exposure, and the clinical presentation varies from single organ involvement (e.g., urticaria, angioedema, rhinitis, conjunctivitis, bronchospasm) to potentially life-threatening anaphylaxis (see figure 2). Mechanistically, most IDHRs rest upon the activation of tissue-resident mast cells and/or circulating basophils. In contrast, NIDHRs occur more than 1 hour after the exposure (often 48-72h later) and mainly manifest as a maculopapular exanthema or, much rarer, as serious cutaneous adverse reactions (SCARs) such as Stevens-Johnson syndrome. These NIDHRs involve the activation of drug-specific T-cells, but not mast cells nor basophils. However, correct allocation of the individual patient to one of these phenotypes can be extremely difficult, if at all possible. Therefore, rapid referral of a patient with a possible drug allergy including a detailed description of the clinical presentation is crucial.

Gathering insights into the underlying pathophysiological mechanism is crucial for correct orientation of further diagnostic work-up of DHRs. A thorough history (and revision of medical records) is of paramount importance. Information about signs (photos when available), symptoms, time of onset of the DHR, treatment of the DHR, index drug, indication for the β -lactam antibiotic, other medication concurrently used, persistence of the signs and symptoms after stopping the medication, re-appearance of the signs and symptoms in absence of the index drug and re-administration of the same drug after the reaction, must be obtained. This information is necessary to differentiate between an IDHR or a NIDHR. Moreover, history (e.g., on the timing of the reaction in response to the last dose of the drug) can be helpful for individual risk stratification. A recent study of our research group showed that an urticarial eruption that appearing within 1 hour after the first intake and regressing within 1 day was significantly more frequently observed in patients with a positive skin test/serum specific IgE assay (1-1-1 criterion) (7).

Figure 2: Signs/symptoms of immediate drug hypersensitivity reactions.

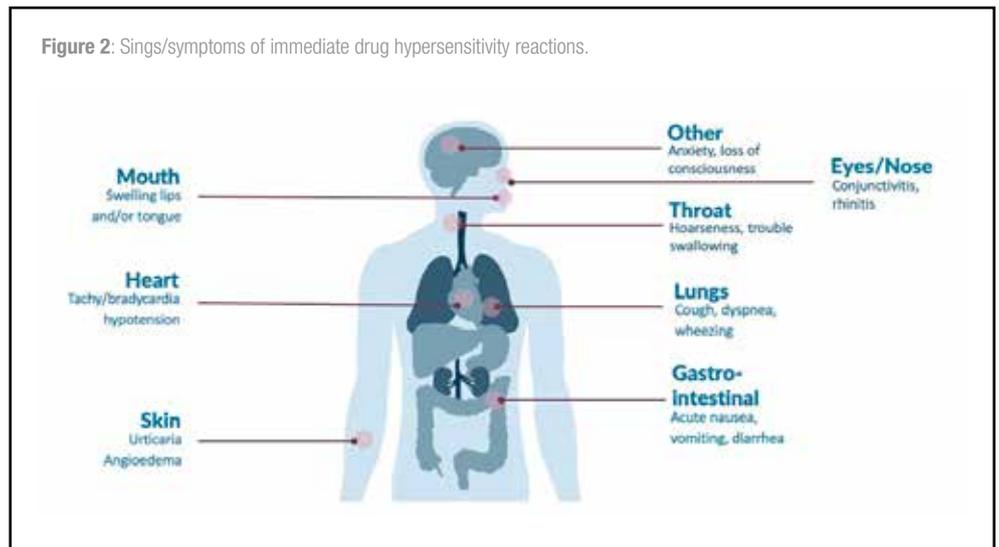
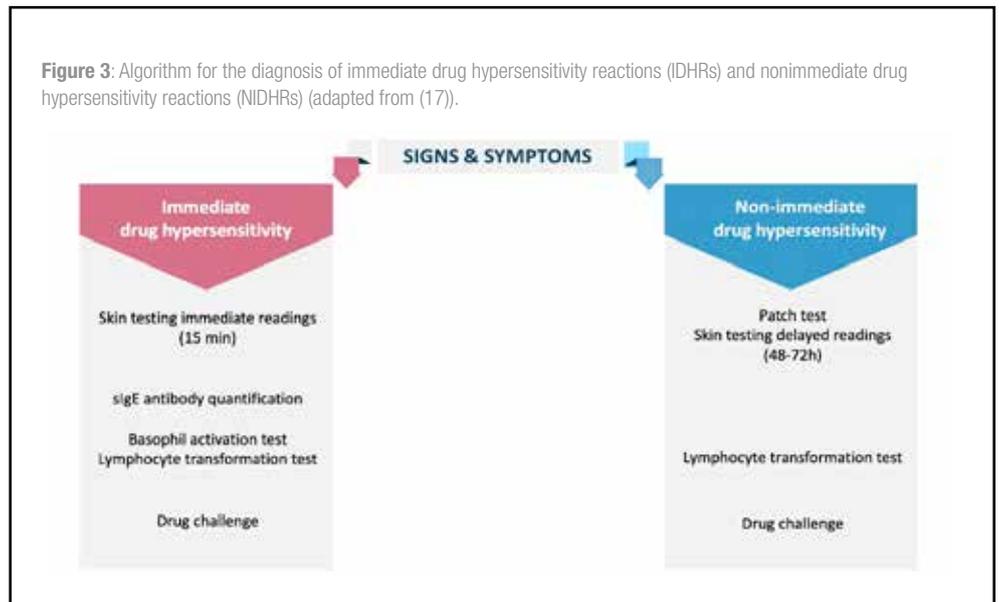


Figure 3: Algorithm for the diagnosis of immediate drug hypersensitivity reactions (IDHRs) and nonimmediate drug hypersensitivity reactions (NIDHRs) (adapted from (17)).



However, the discrimination between IDHR and NIDHR is not always straightforward. Sadly, very often, patients do not remember the exact timing and clinical features of their “index reaction”, as it often occurred decades ago during childhood or adolescence. In these cases, finding out the clinical phenotype is often extremely difficult, if possible at all. In such patients, international guidelines recommend combining diagnostics for immediate and nonimmediate reactions (8). It goes without saying that such a combined approach increases risk and cost. Therefore, quick referral for an allergy workup in case of a possible DHR is recommended.

Confirmatory diagnostics

As mentioned before, further diagnostic work-up is guided by history and the suspected underlying pathophysiological mechanism. Figure 3 shows the diagnostic algorithm of IDHRs and NIDHRs according to the current recommendations (9, 10).

IDHR

Paired serum tryptase

The pathophysiology of IDHRs relies upon the activation and degranulation of mast cells and basophils. Tryptase is a trypsin-like protease that is mainly stored in intracellular mast cell granules. Alfa- and β -protryptase monomers are released spontaneously by resting mast cells. Other α - and β -protryptase monomers are converted

to mature tryptase which is released upon mast cell degranulation. Consequently, an increase in serum tryptase is supportive for mast cell activation.

Thus, paired quantification of acute and basal tryptase is recommended in patients with a suspected IDHR. Acute tryptase should ideally be measured 30 to 120 minutes after the onset of the reaction. Basal tryptase levels should be obtained before the event or at least 24 hours after resolution of all signs and symptoms. The current consensus formula for mast cell activation is a serum acute tryptase level equalling or exceeding 1.2x baseline serum tryptase +2 (11). So even if the value of tryptase at the time of the reaction seems to be normal, that is below 11.4 µg/L, a basal tryptase must be obtained in order to exclude mast cell activation. Noticeably, the absence of mast cell activation does not exclude an IDHR.

Further diagnosis of IDHRs is limited to the demonstration of an IgE-dependent reaction, as no diagnostic is available to demonstrate alternative processes such as mast cell activation via off-target occupation of MRGPRX2 (5). IgE-dependent reactions can be documented *in vitro* by quantification of specific drug-reactive IgE (sIgE) antibodies and *in vivo* skin prick tests and/or intradermal tests with immediate readings.

Skin testing

Today, skin prick testing (SPT) and intradermal testing (IDT), constitute the primary confirmatory step in the diagnostic work-up of an IDHR. Skin prick tests are performed on the ventral part of the forearm and imply a saline buffer solution (negative control) to exclude cutaneous hyperactivity, histamine 10 mg/mL (positive control) to assess skin test reactivity and the involved drugs. SPT are read after 15 minutes and considered positive when the wheal equals or exceeds 3 mm with a surrounding flare. IDT are read after 20 minutes and, for most drugs considered positive when the wheal, accompanied by an erythema, equals or exceeds 5 mm or is doubled as compared to the injection bleb.

Even though skin tests are the first step in the diagnostic work-up of a potential DHR, there are still some disadvantages to be mentioned.

First, skin tests are sometimes unreliable, such as in patients with cutaneous anergy or patients taking antihistamines both leading to false negative readings. False positive results, on the other hand, can be seen in patients with dermatographism. Second, skin testing is not always recommended (e.g., for fluoroquinolones and opiates, skin tests have no added value due to low specificity) (12). Third, skin test performance is highly dependent on the methodology and operator used. Furthermore, for many drugs, the maximal non-irritant concentration (NIC) for skin tests have not been established and have mainly been established in healthy control individuals or have been generalised for all compounds in a certain drug class without correct validation. Besides, NICs might vary for IDHRs and NIDHRs (13). Further validation of NICs is crucial for optimization of sensitivity and specificity of skin tests.

Quantification of total and drug specific IgE antibodies

Quantification of total and drug specific IgE (sIgE) antibodies can be used in the diagnosis of IDHRs. However only a limited number of drug-specific assays are available and sensitivity and specificity of sIgE assays for drugs vary significantly (14). Recently, it has been suggested that, to optimize sensitivity, the threshold for positivity of sIgE has to be lowered to 0.10 kUA/L instead of 0.35 kUA/L. However, a recent study of our research group (15), showed that all patients with a suspected immediate, non-life-threatening, hypersensitivity to amoxicillin or a non-specified penicillin and a sIgE to penicillins between 0.10 kUA/L and 0.35 kUA/L, underwent a drug challenge

without any problems. Diagnosis of penicillin hypersensitivity should not rest upon a low sIgE result alone.

Drug challenges

Drug challenges (DCs) are the reference test to correctly diagnose IDHRs (16). However, DCs entail a risk for (severe) complications and even DCs are not absolutely predictive for the clinical outcome of subsequent exposure with a risk for false negative results (17). Therefore, DCs should be preceded by skin and/or sIgE testing. However, sometimes, in low-risk patients a direct drug challenge can be considered. Traditionally, DCs imply the administration of incremental doses of the suspected drug(s) with a minimum interval of 30 minutes under strict hospital surveillance with emergency room facilities. A minimum observation period of 2 hours after the last dose is recommended. A challenge test is only considered positive when objective symptoms (e.g. hypotension, urticaria, angioedema, wheezing,...) can be observed.

Basophil activation test (BAT), mast cell activation test (MAT) and lymphocyte transformation test (LTT)

As DCs are hampered by the risk of severe, life-threatening reactions and are demanding in resources, time consuming and costly (17), flow-based analyses of basophil activation (BAT) as potential complementary diagnostic for immediate drug allergy have been studied. Although the BAT has become a pervasive test for IDHRs, expert consensus has not been reached. For example, for β-lactam antibiotics, the sensitivity of BAT varies between 13 and a too optimistic 67% (18, 19). The reasons for this poor sensitivity mainly relate to a basophil non-responder status as seen in 10-15% of our patients and rapid negativization of BAT over time (20). Negativization also applies to sIgE and skin tests (21, 22). Importantly, the loss of reactivity in skin testing, quantification of sIgE and BAT, is not necessarily accompanied by loss of clinical reactivity. Whether the passively sensitized mast cell activation test (MAT) might overcome these limitations is a matter of ongoing attractive research. However, because of the rapid decline of sIgE titers, it seems unlikely the MAT will close the gap in the diagnosis of IDHRs to β-lactams. T-cell tests, such as the lymphocyte transformation test (LTT) and variants such as flow-based analysis of activation markers and cytokine expression have only been rarely adopted to document IDHRs (23).

NIDHRs (type IV)

Skin testing

Skin testing procedures for NIDHRs include patch testing and delayed readings of the IDT. Patch testing is a simple and safe diagnostic with a low risk of systemic reactions. In a patch test the drug is generally dissolved in petrolatum and this mixture is applied to the skin of the back. Readings are done after 72-96 hours. Patch testing is considered positive when erythema, infiltration and papules can be observed. Patch testing is the method of choice in patients who experienced SCARs (17). In cases of positive patch tests, further IDTs should be avoided, whereas in cases of negative patch test, IDTs can eventually be performed (9, 10) provided there is no contraindication such as drug-induced autoimmune diseases, severe exfoliative skin reactions and severe vasculitis syndromes.

The technique of IDTs is already described higher. However, for NIDHRs delayed readings of IDT after 48-96 hours are necessary. Delayed readings of IDT are considered positive when an induration surrounded by erythema exceeding 5mm occurs (24). In maculopapular exanthema (MPE), IDTs have a higher sensitivity as compared to patch testing. Therefore, in MPE IDTs are performed without prior patch testing (24).

However, like skin tests for immediate drug allergy, non-irritating concentrations have not yet been established and skin tests are not absolutely predictive. Consequently, again, many patients will need additional DCs to confirm or refute diagnosis.

Drug challenge

As in IDHRs, DC is the reference test for diagnosis of NIDHRs after negative skin testing including delayed readings of IDT and/or patch testing (17). DCs are contraindicated in patients with SCARs and patients with hematologic reactions, e.g. vasculitis (17). As exemplified higher, traditional DCs imply administration of incremental doses of the suspected drug(s) in a single-day. However, for NIDHRs signs and symptoms are expected to occur hours to days after the DC. Prolonged DCs, extending over several consecutive days seem to be of limited use in the diagnosis of nonimmediate drug hypersensitivity (25). It is of utmost importance to balance accuracy, safety, cost, and labor intensity of diagnostic procedures in beta-lactam allergy. Currently, there is increasing evidence for direct challenges in mild NIDHRs. A recent systematic review and meta-analysis (26) showed that in these “low risk” children direct challenges without prior skin testing are effective and safe. However as acknowledged by the EAACI Task Force report (9), hitherto, there is no clear and uniform definition of “mild” and “low-risk”. Further studies on this subject are needed with specific focus on children as their risk profile differs from adults.

Lymphocyte transformation test and variants

There are many in vitro techniques to identify causative agents for NIDHRs such as the LTT, cytokine/mediator detection assays, multiplex bead-based immunoassay and ELISpot. The LTT is the most standardized method but failed to enter mainstream use. The main limitations of the traditional LTT are the need for radioactive thymidine, long culture duration (4-7 days) and poor sensitivity (27). Other in vitro tests, like cytokine detection assays, have also been used, but they are still being evaluated. In the last few years, the advent of performant multicolor flow cytometers has paved the way for the development of novel and more practicable techniques (28). The main advantages of these flow cytometric assays over traditional LTT are speed (48-72h vs. 6 days) and the fact that they do not require a radioisotope. A study of our research group showed that the intracellular quantification of CD154 is an attractive instrument to document both nonimmediate allergies to amoxicillin (clavulanic acid). Most importantly, the test yielded positive results in patients with negative skin tests who needed additional DCs to document diagnosis and it seems that drug-specific T-cell responses might be longer detectable after the index reaction than other diagnostic techniques (29). However, further research is required.

The scourge of unverified “penicillin allergy”

β -lactam antibiotics (β -LABs), especially penicillins, are one of the predominant causes of drug hypersensitivity reactions (DHRs) with significant morbidity and mortality. Alternatively, unverified and false “penicillin allergies”, mainly to the first-line preparations natural penicillin and aminopenicillins, have evolved into a plague with increasing medical and financial burden. A recent study on the prevalence of self-reported penicillin allergy in a Belgian outpatient population showed that 12% of the individuals attending the outpatients’ allergy clinic claimed to have a “penicillin allergy”. However, over 90% of the cases with such a spurious “penicillin allergy” tolerate a challenge with the alleged culprit(s) (30). Importantly, unverified “penicillin allergy” constitutes an almost life-long condition that generally starts as an ill-described, ambiguous and undocumented history in childhood/adolescence going unchallenged in adulthood.

The negative consequences of spurious allergy are undeniable, not

only for the individual patient, but also for society. Actually, spurious “penicillin allergy” is associated with erroneous avoidance and unnecessary substitutions, readmissions, poorer outcomes, prolonged hospitalizations, increased costs and last but not least increased rates of *Clostridium difficile* and antimicrobial resistance. On the other hand, misdiagnosis entails a risk for potentially life-threatening and fatal reactions upon subsequent exposure.

In conclusion, judicious diagnostic work-up by a trained physician is absolutely necessary in every case of both witnessed or self-reported “penicillin allergy”. The importance of a correct and complete description of the index reaction, eventually complemented with pictures of skin lesions, cannot be overemphasized. Every referring physician who witnesses a potential drug hypersensitivity reaction should provide a complete and correct report. This information is critical for guidance of further diagnostic testing and will help to avoid unnecessary tests, which is especially important in children. Further efforts to simplify and optimize the diagnostic approach, to control the plague of alleged “penicillin allergy” and to correctly label patients as truly allergic are needed.

Conflict of interest

The authors have no conflict of interest to declare with regard to the subject discussed in this manuscript.

REFERENCES:

1. Sabato V, Platt P, Garcez T, Cooke P. Suspected perioperative allergic reactions: nomenclature and terminology. *Br J Anaesth*. 2019;123(1):e13-e5.
2. Demoly P, Adkinson NF, Brockow K, Castells M, Chiriac AM, Greenberger PA, et al. International Consensus on drug allergy. *Allergy*. 2014;69(4):420-37.
3. Elst J, Sabato V, Faber MA, Bridts CH, Mertens C, Van Houdt M, et al. MRGPRX2 and Immediate Drug Hypersensitivity: Insights From Cultured Human Mast Cells. *J Investig Allergol Clin Immunol*. 2021;31(6):489-99.
4. Elst J, Maurer M, Sabato V, Faber MA, Bridts CH, Mertens C, et al. Novel Insights on MRGPRX2-Mediated Hypersensitivity to Neuromuscular Blocking Agents And Fluoroquinolones. *Front Immunol*. 2021;12:668962.
5. Sabato V, Ebo DG, Van Der Poorten MM, Toscano A, Van Gasse AL, Mertens C, et al. Allergenic and Mas-Related G Protein-Coupled Receptor X2-Activating Properties of Drugs: Resolving the Two. *J Allergy Clin Immunol Pract*. 2022.
6. Pichler WJ. The p-i Concept: Pharmacological Interaction of Drugs With Immune Receptors. *World Allergy Organ J*. 2008;1(6):96-102.
7. Sabato V, Gaeta F, Valluzzi RL, Van Gasse A, Ebo DG, Romano A. Urticaria: The 1-1-1 Criterion for Optimized Risk Stratification in beta-Lactam Allergy Delabeling. *J Allergy Clin Immunol Pract*. 2021;9(10):3697-704.
8. Dias de Castro E, Carolino F, Carneiro-Leao L, Barbosa J, Ribeiro L, Cernadas JR. Allergy to beta-lactam antibiotics in children: Risk factors for a positive diagnostic work-up. *Allergol Immunopathol (Madr)*. 2020;48(5):417-23.
9. Blanca-Lopez N, Atanaskovic-Markovic M, Gomes ER, Kidon M, Kuyucu S, Mori F, et al. An EAACI Task Force report on allergy to beta-lactams in children: Clinical entities and diagnostic procedures. *Pediatr Allergy Immunol*. 2021;32(7):1426-36.
10. Romano A, Atanaskovic-Markovic M, Barbaud A, Bircher AJ, Brockow K, Caubet JC, et al. Towards a more precise diagnosis of hypersensitivity to beta-lactams - an EAACI position paper. *Allergy*. 2020;75(6):1300-15.
11. Valent P, Bonadonna P, Hartmann K, Broesby-Olsen S, Brockow K, Butterfield JH, et al. Why the 20% + 2 Tryptase Formula Is a Diagnostic Gold Standard for Severe Systemic Mast Cell Activation and Mast Cell Activation Syndrome. *Int Arch Allergy Immunol*. 2019;180(1):44-51.
12. McGee EU, Samuel E, Boronea B, Dillard N, Milby MN, Lewis SJ. Quinolone Allergy. *Pharmacy (Basel)*. 2019;7(3).
13. van der Poorten MM, Van Gasse AL, Hagendorens MM, Faber MA, De Puyssseleyr L, Elst J, et al. Nonirritating skin test concentrations for ceftazidime and aztreonam in patients with a documented beta-lactam allergy. *J Allergy Clin Immunol Pract*. 2021;9(1):585-8 e1.
14. van der Poorten MM, Van Gasse AL, Hagendorens MM, Faber MA, De Puyssseleyr L, Elst J, et al. Serum specific IgE antibodies in immediate drug hypersensitivity. *Clin Chim Acta*. 2020;504:119-24.

15. van der Poorten MM, Van Gasse AL, Hagendorens MM, Faber MA, De Puysseleir L, Elst J, et al. The diagnosis of non-life-threatening immediate penicillin allergy should not rest upon low sIgE results between 0.10 kU/L and 0.35 kU/L in isolation. *Clin Chim Acta*. 2020;511:94-6.
16. Romano A, Valluzzi RL, Gaeta F, Caruso C, Zaffiro A, Quarantino D, et al. The Combined Use of Chronological and Morphological Criteria in the Evaluation of Immediate Penicillin Reactions: Evidence From a Large Study. *J Allergy Clin Immunol Pract*. 2022;10(12):3238-48 e2.
17. Blanca M, Romano A, Torres MJ, Fernandez J, Mayorga C, Rodriguez J, et al. Update on the evaluation of hypersensitivity reactions to betalactams. *Allergy*. 2009;64(2):183-93.
18. Ebo DG, Faber M, Elst J, Van Gasse AL, Bridts CH, Mertens C, et al. In Vitro Diagnosis of Immediate Drug Hypersensitivity During Anesthesia: A Review of the Literature. *J Allergy Clin Immunol Pract*. 2018;6(4):1176-84.
19. Heremans K, Toscano A, Elst J, Van Gasse AL, Mertens C, Beyens M, et al. Basophil Activation Test Shows Poor Sensitivity in Immediate Amoxicillin Allergy. *J Allergy Clin Immunol Pract*. 2022.
20. Ebo DG, Bridts CH, Mertens CH, Sabato V. Principles, potential, and limitations of ex vivo basophil activation by flow cytometry in allergology: A narrative review. *J Allergy Clin Immunol*. 2021;147(4):1143-53.
21. Fernandez TD, Torres MJ, Blanca-Lopez N, Rodriguez-Bada JL, Gomez E, Canto G, et al. Negativization rates of IgE radioimmunoassay and basophil activation test in immediate reactions to penicillins. *Allergy*. 2009;64(2):242-8.
22. Van Gasse AL, Sabato V, Degerbeck F, DeWitt AM, Oulkadi R, Faber MA, et al. Specific IgE to cefazolin: Does it benefit diagnosis? *J Allergy Clin Immunol Pract*. 2019;7(8):2932-4.
23. Glassner A, Dubrall D, Weinhold L, Schmid M, Sachs B. Lymphocyte transformation test for drug allergy detection: When does it work? *Ann Allergy Asthma Immunol*. 2022;129(4):497-506 e3.
24. Romano A, Blanca M, Torres MJ, Bircher A, Aberer W, Brockow K, et al. Diagnosis of nonimmediate reactions to beta-lactam antibiotics. *Allergy*. 2004;59(11):1153-60.
25. Van Gasse AL, Ebo DG, Chiriac AM, Hagendorens MM, Faber MA, Coenen S, et al. The Limited Value of Prolonged Drug Challenges in Nonimmediate Amoxicillin (Clavulanic Acid) Hypersensitivity. *J Allergy Clin Immunol Pract*. 2019;7(7):2225-9 e1.
26. Srisuwatchari W, Phinyo P, Chiriac AM, Saokaew S, Kulalert P. The Safety of the Direct Drug Provocation Test in Beta-Lactam Hypersensitivity in Children: A Systematic Review and Meta-Analysis. *J Allergy Clin Immunol Pract*. 2023;11(2):506-18.
27. Pichler WJ, Tilch J. The lymphocyte transformation test in the diagnosis of drug hypersensitivity. *Allergy*. 2004;59(8):809-20.
28. Ebo DG, Leysen J, Mayorga C, Rozieres A, Knol EF, Terreehorst I. The in vitro diagnosis of drug allergy: status and perspectives. *Allergy*. 2011;66(10):1275-86.
29. Van Gasse AL, Ebo DG, Mertens CM, Bridts CH, Elst J, De Puysseleir L, et al. CD154 (CD40L): A novel aid to document nonimmediate hypersensitivity to amoxicillin or amoxicillin clavulanic acid. *Clin Exp Allergy*. 2020;50(5):640-2.
30. Van Gasse AL, Oulkadi R, Mousati Z, Ebo DG, Chiriac AM, Van Der Poorten MM, et al. Prevalence of self-reported and confirmed penicillin allergy in a Belgian outpatient population. *Allergy*. 2020;75(8):2111-5.